

Seasonal pattern of metal bioaccumulation and their toxicity on *Sphagnum squarrosum*

Anuj Saxena

Department of Botany, Sacred Heart Degree College, Naipalapur, Sitapur-261 001, India

(Received: 8 April, 2004 ; Accepted: 22 January, 2005)

Abstract : Present study was undertaken as an attempt to study the effect of pollutants on biological responses of *Sphagnum* growing at Kainchi, Kumaon hills (Utranchal). *Sphagnum* plants of almost identical size, collected from the marked sites of Kainchi in different seasons viz., monsoon, winter, summer and again in monsoon, were analysed for chlorophyll, protein, shoot length and nitrate reductase and peroxidase activities. Maximum chlorophyll, protein, shoots length and nitrate reductase activities were observed during the monsoon while minimum in summers. The abundance of *Sphagnum* and two other bryophytes, *Marchantia* and *Plagiochasma* was also higher in monsoon than in other seasons. The study also indicated that *Sphagnum* has more bioaccumulation and tolerance potential for heavy metals than *Marchantia* and *Plagiochasma*.

Key words : Bryophytes, Metal pollution, Phytotoxicity

Introduction

Bryophytes particularly the mosses have been known to accumulate atmospheric particulates and metals from dry and wet depositions (Weiss *et al.*, 1997; Vasconcelos and Taveres, 1998). This has made them popular for use in biomonitoring and phytoremediation studies of air pollutants. Valuable knowledge about input of anthropogenic metals from local and distinct emission sources could be obtained from the physiological studies and chemical analysis of mosses (Syso, 1998). *Sphagnum* moss has been used in pollution studies on several occasions (Glooschenko and Capobianco, 1976; Ho *et al.*, 1996a; Saxena, 1999) and metals have found to be accumulated readily by the shoots mainly by cation exchange at cell walls (Ho *et al.*, 1996b; Saxena *et al.*, 2003).

The suitability of mosses particularly *Sphagnum* as bioindicator is mainly due to one cell thick leaf structure, lack of any protective layer like cuticle or epidermis, absorption and accumulation of nutrients or pollutants directly from the atmosphere, absence of vascular supply for transportation of minerals within the plant, green and perennial growth in wide variety of habitat, ability to be stored for several years without any special care and easy chemical analysis (Callghan *et al.*, 1978; Baker, 1992).

The pollutants may have indirect effect on population caused by alteration in the habitat, which in turn is an indicator of intensity of pollutants in environment or in ecosystem that can be assessed by change in community structure or growth. Growth of an organism can be considered to be the integrated result of numerous physiological and biochemical responses of environmental parameters and therefore, it can also be used to judge the intensity of pollution.

Materials and Methods

Study sites: Kainchi (District Nainital) is located at an altitude of 1873 msl. (5900 feet) on Kumaon hills, situated along the

road no. 37, connecting Almora and Ranikhet from Bareilly via Haldwani. Its climate varies through out the year. It is quite cold in winter and mild warm through May-June followed by monsoon rains till October. The average rainfall is 200 cm; minimum and maximum temperatures are 4 °C and 36 °C; relative humidity 60-85%, highest in the month of July-August and lowest in January.

Sampling and preparation of moss bags: *Sphagnum squarrosum* Crome. Samml plants were collected from undisturbed habitat of Muketswar (Kumaon hills of Utranchal) located at 7500 feet and brought to the laboratory in polythene bags. Plants were washed several times and sorted for gametophytes only. Nine moss bags were prepared for transplantation at three sites in triplicate. Each nylon bag was of 1.8" X 1.8" quadrangular plastic frame, interwoven with polymer wire net. 20 gm (dry wt.) of *Sphagnum* was pressed between the folds of moss bag. Moss bags were transplanted at catchment sites for three successive periods (each of four months).

Biological studies: *Sphagnum squarrosum*, *Marchantia polymorpha* and *Plagiochasma appendiculatum* collected from Mukteswar and Kainchi in different seasons were analysed for biochemical and physiological parameters. *In vivo* nitrate reductase activity was measured in accordance to the method of Srivastava (1975) by spectrophotometrically quantifying the nitrite released into the incubation medium. The colour is developed due to the formation of diazo compound with sulphanilamide and nitrite, which is coupled with NED. For peroxidase estimation the method of Putter (1974) was followed using guaiacol as a dye. Protein was estimated following the Folin-phenol method of Lowry *et al.* (1951) using Bovin Serum albumin as standard. Chlorophyll was estimated according to modified Arnon (1949) method by extracting the pigment in 80% acetone. Method of Shimwell and Laurie (1972) was adopted for metal analysis by digesting 1 g of oven dried material (80 ± 2

°C for 6 hrs) in concentrated (3 : 1) HClO₄ : HNO₃. The metal content was estimated by atomic absorption spectrophotometer (Perkin-Elmer model).

The data presented are average of three replicates with SE. For statistical analysis of data Dunken Multiple Range Test (mean separation test) was applied.

Results and Discussion

Seasonal oscillation in biological parameters of *Sphagnum*: Higher chlorophyll, total protein, dry matter, shoot density, shoot length and nitrate reductase activity were recorded during the monsoon followed by winter, whereas a sharp decline in above parameters was observed during the summer in the moss samples collected from Kainchi. However, peroxidase activity followed a reverse trend (Table 1). A very minor seasonal variation was observed in above parameters in the *Sphagnum* collected from Mukteswar (Table 2).

Experiments demonstrate that the October is the best month for the overall growth and metabolic state of *Sphagnum* and other bryophytes. Low leaf area indices were observed during summers (data not shown), which is particularly due to low shoot density or number of leaves per unit area. The shoot density depends upon light, water and growing seasons

(Proctor, 1982; Bates, 1988; Van der Hoeven and During, 1997). But here seasonal variations in shoot density and leaf area indices were observed, which is strongly correlated with seasons and metal accumulation pattern. This needs a further investigated.

Certain physiological and biochemical effects of metals on higher plants have been reviewed by several workers (Thapa *et al.*, 1988; Pahlsson, 1989). Decline in chlorophyll content has been shown to be a reliable criterion in many studies with environmental stresses, including heavy metals. Metal induced decline in chlorophyll content has been demonstrated in some higher plants (Burzynski, 1985; Sinha *et al.*, 1988). The decline is apparently due to reduced chlorophyll synthesis (Nag *et al.*, 1981), which is the consequence of interaction of metal with some vital enzymes involved in the synthesis of chlorophyll (Stobart *et al.*, 1985). The decreased chlorophyll content may lead to reduce photosynthesis and ultimately to reduce biomass. The present study suggests that seasonal variation in chlorophyll content can be correlated with metal precipitation pattern of atmosphere.

Inhibition of nitrogenous parameters such as protein, total nitrogen and nitrate reductase activity under metal stress has been observed in many higher plants (Bhandal and Kaur,

Table – 1: Physiological parameters of *Sphagnum* harvested in different seasons from marked sites of Kainchi.

Parameters	Seasons			
	October, 1997 (Monsoon)	February, 1998 (Winter)	June, 1998 (Summer)	October, 1998 (Monsoon)
Chlorophyll (mg g ⁻¹ FW)	3.08 ± 0.07 ^a	2.87 ± 0.09 ^a	2.33 ± 0.06 ^b	2.95 ± 0.07 ^a
Nitrate reductase (n mol NO ₂ h ⁻¹ g ⁻¹ FW)	5017 ± 3.37 ^a	4968 ± 5.29 ^a	4266 ± 2.56 ^b	5021 ± 3.47 ^a
Protein (mg g ⁻¹ FW)	17.64 ± 0.13 ^a	16.85 ± 0.11 ^b	13.92 ± 0.13 ^c	17.58 ± 0.08 ^a
Peroxidase (∪ OD min ⁻¹ g ⁻¹ FW)	0.185 ± 0.06 ^a	0.265 ± 0.07 ^b	0.495 ± 0.09 ^c	0.197 ± 0.05 ^a
Moisture content (percent)	68 ± 1.81 ^a	65 ± 0.74 ^a	59 ± 0.88 ^b	66 ± 0.05 ^a
Dry matter (g cm ⁻²)	0.37 ± 0.02 ^a	0.29 ± 0.01 ^b	0.13 ± 0.01 ^c	0.38 ± 0.02 ^a
Shoot length (mm)	65 ± 2.0 ^a	56 ± 2.08 ^b	37 ± 2.0 ^c	67 ± 1.73 ^a
Dry weight (mg per plant)	8.0 ± 0.15 ^a	7.6 ± 0.20 ^a	4.6 ± 0.11 ^b	7.8 ± 0.21 ^a

**Sphagnum* samples were analysed in the laboratory within 24 hrs of harvesting from their natural habitat (Kainchi). Values with the same superscript are statistically same.

Table – 2: Physiological parameters of *Sphagnum* harvested in different seasons from marked sites of Mukteswar.

Parameters	Seasons			
	October, 1997 (Monsoon)	February, 1998 (Winter)	June, 1998 (Summer)	October, 1998 (Monsoon)
Chlorophyll (mg g ⁻¹ FW)	3.32 ± 0.05 ^a	3.27 ± 0.02 ^a	3.18 ± 0.02 ^a	3.29 ± 0.02 ^a
Nitrate reductase (n mol NO ₂ h ⁻¹ g ⁻¹ FW)	5056 ± 3.87 ^a	4982 ± 2.20 ^b	5036 ± 2.53 ^{a,b}	5066 ± 4.67 ^a
Protein (mg g ⁻¹ FW)	18.24 ± 0.07 ^a	17.71 ± 0.02 ^b	15.51 ± 0.03 ^c	17.86 ± 0.03 ^b
Peroxidase (∪ OD min ⁻¹ g ⁻¹ FW)	0.135 ± 0.07 ^a	0.124 ± 0.01 ^b	0.138 ± 0.01 ^c	0.115 ± 0.01 ^b
Moisture content (percent)	70 ± 0.30 ^a	67 ± 0.19 ^b	61 ± 0.15 ^c	69 ± 0.30 ^a
Dry matter (g cm ⁻²)	0.42 ± 0.02 ^a	0.39 ± 0.02 ^a	0.36 ± 0.01 ^b	0.41 ± 0.02 ^a
Shoot length (mm)	68.8 ± 0.32 ^a	61.93 ± 0.32 ^b	60.13 ± 0.12 ^c	70.73 ± 0.06 ^a
Dry weight (mg per plant)	8.3 ± 0.09 ^a	7.9 ± 0.04 ^a	7.6 ± 0.06 ^b	8.1 ± 0.08 ^{a,b}

**Sphagnum* samples were analysed in the laboratory within 24 hrs of harvesting from their natural habitat (Mukteswar). Values with the same superscript are statistically same.

Table – 3: Distribution of bryophytes in natural habitat at Kainchi and Mukteswar in different seasons.

Seasons	Species	Abundance*	
		Kainchi	Mukteswar
March to June (Summer)	<i>Sphagnum sp.</i>	+++	++
	<i>Marchantia sp.</i>	+	+++
	<i>Plagiochasma sp.</i>	++	++++
July to October (Monsoon)	<i>Sphagnum sp.</i>	++++	+++
	<i>Marchantia sp.</i>	+++	+++++
	<i>Plagiochasma sp.</i>	++++	++++++
November to February (Winter)	<i>Sphagnum sp.</i>	++++	+++
	<i>Marchantia sp.</i>	+++	+++++
	<i>Plagiochasma sp.</i>	+++	+++++

* Each + mark represent population at three sites.

Table – 4: Determination of metal content ($\mu\text{g g}^{-1}$ dw) at Kainchi during 1998-99 in native bryophytes.

Seasons	Species	Pb	Fe	Cu	Cd	Ni	Zn	Total
June	<i>S. squarrosum</i>	32.83 ± 1.73	91.3 ± 1.86	6.6 ± 0.28	0.63 ± 0.03	5.4 ± 0.02	25.7 ± 1.33	162.46 ± 3.22
	<i>M. polymorpha</i>	14.17 ± 0.36	41.38 ± 0.53	2.84 ± 0.13	0.23 ± 0.01	2.59 ± 0.06	11.58 ± 0.12	72.79 ± 1.86
	<i>P. appendiculatum</i>	21.84 ± 0.48	48.18 ± 0.79	4.09 ± 0.21	0.38 ± 0.02	3.79 ± 0.08	16.31 ± 0.83	94.59 ± 1.79
October	<i>S. squarrosum</i>	18.37 ± 0.27	38.24 ± 0.58	3.37 ± 0.24	0.36 ± 0.02	3.36 ± 0.13	22.47 ± 1.37	86.17 ± 0.96
	<i>M. polymorpha</i>	8.84 ± 0.08	9.47 ± 0.65	1.97 ± 0.16	9.13 ± 0.01	1.41 ± 0.09	7.67 ± 0.09	38.49 ± 0.74
	<i>P. appendiculatum</i>	11.39 ± 0.12	17.76 ± 0.39	2.89 ± 0.17	0.24 ± 0.01	2.27 ± 0.11	11.73 ± 0.04	46.28 ± 0.93
February	<i>S. squarrosum</i>	16.64 ± 0.86	42.37 ± 1.16	5.14 ± 0.23	9.28 ± 0.30	2.95 ± 0.13	28.86 ± 0.67	105.24 ± 0.59
	<i>M. polymorpha</i>	7.48 ± 0.32	15.91 ± 0.67	1.87 ± 0.15	9.10 ± 0.13	1.03 ± 0.09	7.49 ± 0.38	42.88 ± 0.87
	<i>P. appendiculatum</i>	10.75 ± 0.74	26.68 ± 0.72	3.28 ± 0.12	0.17 ± 0.01	1.67 ± 0.08	12.63 ± 0.76	55.18 ± 1.36

Table – 5: Determination of metal content ($\mu\text{g g}^{-1}$ dw) at Mukteswar during 1998-99 in native bryophytes.

Seasons	Species	Pb	Fe	Cu	Cd	Ni	Zn	Total
June	<i>S. squarrosum</i>	7.46 ± 0.87	43.05 ± 1.35	3.97 ± 0.48	0.37 ± 0.04	1.15 ± 0.19	11.37 ± 0.81	66.32 ± 0.94
	<i>M. polymorpha</i>	3.14 ± 0.53	24.41 ± 0.87	1.26 ± 0.19	0.14 ± 0.01	0.41 ± 0.09	5.49 ± 0.32	34.84 ± 0.53
	<i>P. appendiculatum</i>	4.97 ± 0.48	32.74 ± 0.76	2.28 ± 0.23	0.17 ± 0.02	0.76 ± 0.12	7.31 ± 0.43	48.25 ± 0.37
October	<i>S. squarrosum</i>	6.63 ± 0.47	34.13 ± 0.52	3.15 ± 0.47	0.27 ± 0.02	0.83 ± 0.14	13.63 ± 0.57	60.60 ± 0.47
	<i>M. polymorpha</i>	2.47 ± 0.47	19.38 ± 0.43	1.73 ± 0.39	0.11 ± 0.01	0.31 ± 0.13	6.41 ± 0.09	30.69 ± 0.23
	<i>P. appendiculatum</i>	3.98 ± 0.51	23.89 ± 0.27	2.14 ± 0.17	0.23 ± 0.01	0.59 ± 0.08	9.53 ± 0.14	40.55 ± 0.39
February	<i>S. squarrosum</i>	6.59 ± 0.43	39.46 ± 0.56	2.93 ± 0.43	0.31 ± 0.02	0.74 ± 0.18	16.57 ± 0.48	66.52 ± 0.57
	<i>M. polymorpha</i>	2.75 ± 0.34	19.78 ± 0.32	1.21 ± 0.54	0.11 ± 0.01	0.27 ± 0.36	6.97 ± 0.53	30.79 ± 0.38
	<i>P. appendiculatum</i>	4.17 ± 0.56	25.17 ± 0.45	1.87 ± 0.32	0.19 ± 0.01	0.53 ± 0.43	10.58 ± 0.34	42.26 ± 0.43

1992; Kevresan *et al.*, 1998). Inhibition of NR activity in *Sphagnum* has been demonstrated in earlier study also which is apparently due to the interaction of the metal with –SH groups of enzyme (Saxena *et al.*, 1999).

Variation in peroxidase activity may be due to seasonal variation in metal accumulation by *Sphagnum*, as reported here. It might be an indication of the induction of free radical scavenging metabolism, which is often seen under different environmental stresses (Van Assche and Clijster, 1990). Increased peroxidase activity has also reported in response to metals in barley and in sunflower (Gallego *et al.*, 1996; Jita *et al.*, 1998).

Seasonal variation in abundance of bryophytes: Although *Sphagnum* is more frequently distributed at Kainchi than at

Mukteswar, its abundance varies from season to season at Kainchi. Whereas at Mukteswar *Sphagnum* is more or less uniformly distributed throughout the year. Seasonal variation in abundance of *Marchantia* and *Plagiochasma* was also observed specially at Kainchi (Table 3).

Metal content in native and transplanted bryophytes: A clear-cut difference was observed in metal content of all three studied bryophytes harvested from Kainchi and Mukteswar in different seasons. Mukteswar represented a pure, undisturbed and ideal niche for the luxuriant growth of most of the bryophytes as very low level of metals was detected with a very insignificant seasonal variation. (Table 4, 5 and 6). It was however observed that overall metal concentration was higher during March to June (*i.e.* summers), while lowest during July

Table – 6: Determination of metal content ($\mu\text{g g}^{-1}$ dw) at Kainchi during 1998-99 in *Sphagnum squarrosum* transplanted at Kainchi.

Seasons	Site	Pb	Fe	Cu	Cd	Ni	Zn	Total
Pretransplanted <i>Sphagnum</i>		6.63 \pm 0.32	39.46 \pm 0.56	2.93 \pm 0.30	0.19 \pm 0.03	0.74 \pm 0.03	16.57 \pm 0.17	66.52 \pm 1.21
July to	Post office	31.47 \pm 0.47	138.01 \pm 0.96	18.31 \pm 0.8	0.56 \pm 0.04	3.43 \pm 0.93	54.63 \pm 0.91	246.41 \pm 3.17
October	Plantiss agrotech	15.93 \pm 0.57	94.16 \pm 1.53	26.83 \pm 1.21	0.34 \pm 0.02	2.17 \pm 0.85	69.51 \pm 1.17	208.94 \pm 3.43
	Kainchi temple	24.56 \pm 0.75	119.17 \pm 2.63	20.41 \pm 0.52	0.45 \pm 0.03	3.23 \pm 0.77	48.47 \pm 0.81	216.29 \pm 4.22
November to	Post office	38.12 \pm 0.39	131.95 \pm 2.91	27.63 \pm 0.73	0.36 \pm 0.02	1.93 \pm 0.53	69.21 \pm 1.22	269.20 \pm 1.97
February	Plantiss agrotech	18.57 \pm 0.34	119.80 \pm 1.18	27.63 \pm 0.61	0.29 \pm 0.01	1.93 \pm 0.87	83.65 \pm 1.05	251.87 \pm 1.34
	Kainchi temple	31.79 \pm 0.53	186.35 \pm 2.54	16.64 \pm 0.47	0.31 \pm 0.02	2.27 \pm 0.59	61.35 \pm 2.36	298.71 \pm 1.62
March to	Post office	69.41 \pm 1.21	212.58 \pm 3.87	15.92 \pm 0.9	0.93 \pm 0.04	4.36 \pm 0.78	61.35 \pm 0.57	364.55 \pm 3.67
June	Plantiss agrotech	38.84 \pm 0.67	91.21 \pm 1.14	18.43 \pm 1.13	0.66 \pm 0.02	3.18 \pm 0.94	86.07 \pm 1.87	238.39 \pm 2.94
	Kainchi temple	55.83 \pm 0.78	116.39 \pm 1.11	14.37 \pm 0.53	0.81 \pm 0.05	3.9 \pm 1.07	52.24 \pm 0.96	243.54 \pm 2.72

to October. Lower value of metals could be attributed to the (i) decrease in the amount of metal available due to lower number of tourist during rainy seasons, (ii) leaching of pollutants with rain water, (iii) increase in biomass and growth more rapidly in rains due to favorable conditions that might decrease the percentage of metal in proportion to biomass.

The study fairly demonstrates that moss *Sphagnum* can accumulate and tolerate elevated levels of metals without showing any toxic symptoms. Therefore, it can be used as an ideal tool for biomonitoring and phytoremediation studies.

Acknowledgements

Author wish to extend his sincere thanks to Prof. Janice M. Glime, Michigan Technological University, Houghton for critical suggestions from time to time and to late Prof. H. S. Srivastava, Department of Plant Science, M. J. P. Rohilkhand University, Bareilly for encouragement.

References

- Amon, D. I.: Copper enzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris*. *Plant Physiol.*, **24**, 1-15 (1949).
- Baker, R. G. E.: The influence of copper and zinc on shoot length and dry weight of *Sphagnum palustre* and *Sphagnum cuspidatum* in aqueous cultures. *J. Hattori Bot. Lab.*, **72**, 89-96 (1992).
- Bates, J. W.: The effect of shoot spacing on the growth and branch development of moss *Rhytidiadelphus triquetrus*. *New Phytol.*, **1**, 499-504 (1988).
- Bhandal, I. S. and H. Kaur: Heavy metal inhibition of nitrate uptake and *in vivo* nitrate reductase in roots of wheat, *Triticum aestivum*. *Ind. J. Plant Physiol.* **35**, 281-284 (1992).
- Burzynski, M.: influence of lead on the chlorophyll content of initial steps on its synthesis in greening cucumber seedlings. *Acta Bot. Soc. Bot. Poll.*, **54**, 95-105 (1985).
- Callghan, T. V., N. I. Collins and C. H. Callaghan: Photosynthesis, growth and reproduction of *Hylocomium splendens* and *Polytrichum commune* in Swedish Lapland. Strategies of growth and population dynamics of tundra plants. *Oikos*, **3**, 73-88 (1978).
- Gallego, S. M., M. P. Benavides and M. L. Tomaro: Oxidative damage caused by cadmium chloride in sunflower (*Helianthus annuus*). *Plant Physiol.*, **58**, 41-52. (1996).
- Glooschenko, W. A. and J. A. Capobianco: Metal content of the *Sphagnum* mosses from two northern Canadian bog ecosystems. *Water, Air and Soil Pollut.*, **10**, 215-220 (1976).
- Ho, Y. S., D. A. J. Wase, and C. F. Forster: Kinetic studies of competitive heavy metal absorption by *Sphagnum* moss peat. *Environ. Technol.*, **17**(1), 71-77 (1996a).
- Ho, Y. S., D. A. J. Wase, and C. F. Forster: Removal of lead ions from aqueous solution using *Sphagnum* moss peat as adsorbant. *Water- SA*, **22**(3), 219-224 (1996b).
- Jita, P., B. B. Panda and J. Pantra: A comparison of biochemical responses to oxidative and metal stress in seedling of barley, *Hordeum vulgare* L. *Environ. Pollut.* **101**, 99-105 (1998).
- Kevresan, S., M. Papovie, J. Kandrac and N. Petrovie: Effect of heavy metals on nitrate and protein metabolism in sugar beat. *Biologia Plantarum* (Czech Republic), **41**(2), 235-240 (1998).
- Lowry, O. H., N. J. Rose, A. L. Brough and N. R. J. Randall: Protein measurement with folin phenol reagent. *J. Biol. Chem.*, **193**, 265-275 (1951).
- Nag, P., A. K. Paul and S. K. Mukherji: Heavy metal effects in plant tissues involving chlorophyll, chlorophyllase, Hill reaction activity and gel electrophoretic patterns of soluble proteins. *Ind. J. Exp. Biol.*, **19**, 702-706 (1981).
- Pahlsson, A. B.: Toxicity of heavy metals (Zn, Cu, Cd, Pb) to vascular plants. *Air & Soil Pollut.*, **47**, 287-319 (1989).
- Proctor, M. C. F.: Physiological ecology: water relations, light and temperature responses, carbon balance. *In* : Bryophyte Ecology (Ed: I. Smith). Chapman and Hall, Cambridge. pp. 333-382. (1982)
- Putter, J.: *In*: Methods of enzymatic analysis 2 (Ed: Bergmeyer). Academic Press, New York pp. 685 (1974).
- Saxena, D. K.: Biomonitoring of Pb, Ni, Cr, Hg with the help of bryophytes in Nainital. *In* : Proceeding of National Conference on Bryology, (Ed: V. Nath, Bishen Singh Mahendra Pal Singh) pp.159-176 (1999).
- Saxena, D. K., A. Saxena, and H. S. Srivastava: Heavy metal accumulation and *in vivo* nitrate reductase activity in the *Sphagnum squarrosum* Cram. *Samml. Proc. Nat. Acad. Sci.*, **69B** (III & IV), 307-312 (1999).
- Saxena, A., D. K. Saxena and H. S. Srivastava: The influence of glutathione on physiological effects of lead and its accumulation in moss *Sphagnum squarrosum*. *Water, Air and Soil Pollut.*, **143**, 351-361 (2003).
- Shimwell, D. W. and A. J. Laurie: Lead and zinc contamination of vegetation in the Southern Pennines. *Environ. Pollut.*, **3**, 291-301 (1972).

- Sinha, S. K., H. S. Srivastava and S. N. Mishra: Nitrate assimilation in intact and excised maize leaves in the presence of lead. *Bull. Environ. Contam. Toxicol.*, **41**, 419-426 (1988).
- Srivastava, H. S.: Distribution of nitrate reductase in ageing bean seedlings. *Plant & Cell Physiol.*, **16**, 995-999 (1975).
- Stobart, A. K., W. T. Griffith, I. Ammen-Bukhari and R. P. Sherwood: The effect of Cd²⁺ on the biosynthesis of chlorophyll in leaves of barley. *Plant Physiol.*, **63**, 293-298 (1985).
- Syso, A. S.: Using the Cr, Ni relationship for monitoring environmental pollution. *Agrokhimya*, **4**, 76-83 (1998).
- Thapa, D., H. S. Srivastava and D. I. Ormnod: Physiological and biochemical effects of lead on higher plants. *Vegetos*, **1**, 107-119 (1988).
- Van Assche, F. and J. Clijster: Effects of metals on enzyme activity of plants. *Plant Cell Environ.*, **19**, 195-206 (1990).
- Van der Hoeven, E. C. and H. J. During: The effect of density on size frequency distributions in chalk grassland bryophyte populations. *Oikos*, **80**, 533-539 (1997).
- Vasconcelos, M. T. and H. M. Taveres: Atmospheric metal pollution (Cr, Cu, Fe, Mn, Ni, Pb and Zn) in Oporto city derived from results for low volume aerosol sampler and for the moss *Sphagnum auriculatum* bioindicator. *Sci. Total Environ.*, **212**, 11-29 (1998).
- Weiss, D., W. Shotyk, A. K. Cheburkin, M. Gloor and S. Reese: Atmospheric lead deposition from 12400 to Ca2000 yrs BC in a peat bog profile, in Jura mountains Switzerland. *Water, Air and Soil Pollut.*, **100**, 311-324 (1997).

Correspondence to :

Dr. Anuj Saxena

Department of Botany,

Sacred Heart Degree College,

Sitapur-261 001 (U.P.), India

E-mail: anujsaxena011@rediffmail.com